Stabilization of Returned Dairy Products by Ensiling with Straw and Molasses for Animal Feeding

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ABSTRACT

Returned dairy products which are transferred to landfills might add to the environmental pollution. Such products have a high nutritional value for ruminants, but they should be stabilized to enable their use as cattle feed. The purpose of the current study was to examine stabilization of returned dairy products by ensiling in combinations with straw and molasses for animal feeding. Treatments included combinations of milk and cottage cheese with straw and molasses. Results indicate that such products ensile well with straw, and after 3 d of ensiling the pH decreased to around 4.0. It was necessary to supplement cottage cheese with molasses, to supply a carbohydrate source for the lactic acid fermentation. The major fermentation product was lactic acid. Percentage of ammonia N (of total N) was generally higher in the silages made with cottage cheese than in those made with milk; the highest percentage (16%) was measured in the second experiment in the silages prepared with cottage cheese and straw. The study indicates the potential of stabilizing returned dairy products for animal feeding along with straw and molasses. There may also be potential for large dairy farms, or groups of smaller farms, to ensile waste milk with straw for later use as feed.

(Key words: dairy products, ensiling, cattle feeding)

Abbreviation key: WSC = water soluble carbohydrates.

INTRODUCTION

Many dairy products are returned from the grocery stores because the sale-by date has been reached or because of spoilage, which is enhanced in warm climate, in spite of cooling systems. In Israel out of the billionliter market of dairy products about 2% are returned. These include milk, soft cheeses, and cream. The returned dairy products are transferred along with municipal trash to landfills and contribute to the environmental load. Effluent from spoiling dairy products might leak into underground water sources and result in pollution. The biological oxygen demand values for wastewater generated in the dairy industry are 500 to 700 and 1500 to 2000 ppm O_2 , for milk and cheese, respectively (Wheatley, 1994). The values for the actua' dairy products might be much higher.

Milk and dairy products have a high nutritional value for animals due to their protein, fat, sugar, and mineral content. Therefore, it would be beneficial to utilize returns of such products for animal feeding. All products are originally prepared from pasteurized milk and therefore, the potential health hazards to animals consuming such products are minimal. However, for efficient utilization the returned dairy products should be stabilized to enable short-term storage and handling. Stabilization of returned products should be cost efficient. One possibility for preservation is ensiling which is a preservation method for moist crops. Ensiling is based on spontaneous solid-state fermentation whereby lactic acid bacteria convert water-soluble carbohydrates into organic acids, mainly lactic acid. As a result the pH decreases, and the forage is preserved. Returned dairy products are too moist for ensiling because of potential fermentation and effluent problems. Therefore, it would be desirable to mix them with a dry mate rial to absorb excess moisture. Sugar addition may 🥿 necessary to adequate fermentation.

The purpose of the current work was to study the ensiling properties of dairy products in combination with straw and molasses.

MATERIALS AND METHODS

Fresh homogenized milk (3% fat) and cottage cheese (5% fat) packed in 2 kg plastic bags were donated by a local dairy plant. Two ensiling experiments were performed: immediately upon receipt and 1 wk later on the sale-by date. The products were stored at 4° C until used.

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Table 1. Analysis of the fresh materials. Lactobacilli are given as \log_{10} number of colony forming units g⁻¹.

	DM, g/kg	рН		WSC ² , g/kg		Lactobacilli	
		Exp. 1	Exp. 2	Exp. 1	Exp. 2	Exp. 1	Exp. 2
Straw ¹	862	3	6.7		24	3	4.6
Cottage cheese	198	5.2	5.5	9	0	6.4	6.8
Milk	115	7.0	6.8	33	38	<2.0	5.8

 $^{1}\mathrm{Rumen}$ degradability of the straw 605 \pm 21 g/kg; no yeasts or molds were detected in the milk or in the cottage cheese.

²WSC = water-soluble carbohydrates.

³Not measured.

Treatments were comprised of mixtures of dairy products and wheat straw to obtain adequate DM composition for ensiling (approximately 350 g/kg), with or without molasses as a carbohydrate source. The same wheat straw and molasses were used in both experiments.

Experiment 1

The following treatments were prepared in triplicates: 1) 830 g cottage cheese mixed with 75 g chopped wheat straw, 2) treatment 1 plus 26 g molasses, 3) 700 g milk mixed with 265 g chopped straw, and 4) 465 g of treatment 3 mixture plus 15 g molasses. The various mixtures were ensiled in 0.25-L sealed jars and stored for 30 d at 30°C. The wet weights were 185 ± 7 and 118 ± 2 g per jar, respectively, for the cottage cheese and straw and for the milk and straw mixtures, respectively.

Experiment 2

The treatments were prepared as follows: 1) 3 kg of cottage cheese were mixed with 0.75 kg of chopped straw, 2) one-half of treatment 1 plus 50 g molasses, 3) 2.85 kg milk were mixed with 1.15 kg of chopped straw, and 4) one-half of treatment 3 plus 50 g molasses. The various mixtures were ensiled in 0.25-L sealed jars

and stored at 30°C. There were nine jars per treatment, and three were opened on d 3, 7, and 36 to follow fermentation dynamics. The wet weights were 110 ± 8 and 134 ± 7 g per jar for the cottage cheese and straw and for the milk and straw mixtures, respectively.

Analyses

Chemical analyses of the raw materials and fresh mixtures were conducted on pooled single samples, and analyses of the silages were performed on individual jars (triplicates). Dry matter was determined by oven drying for 48 h at 60°C. Crude protein was determined by the Kjeldahl method (AOAC, 1980). Ammonia N was determined by the Kjeldahl method without the digestion step as follows: a 40 g sample was extracted in 360 ml distilled water in a Stomacher blender for 3 min. The extract was filtered through Whatman No 1 paper, and 100 ml was processed in a Kjeltech auto 1030 analyzer (Tecator, Sweden). Water-soluble carbohydrates (WSC) were determined by the phenol-sulfuric acid method according to Dubois et al. (1956). Lactic acid was determined by a spectrophotometric method according to Barker and Summerson (1941). Volatile fermentation products were determined in aqueous extracts by means of a GLC with a semi-capillary FFAP (nitroterephthalic acid-modified polyethylene glycol)

Table 2. Chemical analysis of the ensiled dairy products after 30 d of ensiling—Experiment 1. Results are in g/kg of DM, except as noted.

Treatment ¹	DM g/kg	pH	WSC ⁴	Lactic acid	Acetic acid	Ethanol	Crude protein	NH ₃ -N%/TN ⁴	RD^4
1^{2}	303	5.743	10^{c3}	6 ^{c3}	1843	5 ⁴³	394 ³	13.3	710 ^{b3}
2	307	4.9^{b}	6°	$27^{\rm b}$	$19^{\rm a}$	6 ^a	$381^{\rm b}$	7 ^b	799
3	327	3.9°	26^{b}	49^{a}	$10^{\rm b}$	Ob	130 ^c	2^{c}	617°
4	341	3.9°	46 ^a	45^{a}	8 ^b	O _j ,	128°	1^{c}	$707^{\rm b}$

¹Treatments: 1) cottage cheese and straw; 2) cottage cheese, straw, and molasses; 3) milk and straw; 4) milk, straw, and molasses.

 $^2\mathrm{Treatment}$ 1 contained also 5 g kg 1 DM but yric acid.

 3 Within a column, means followed by different superscripts are significanly different (P < 0.05).

 $^{4}\mathrm{WSC}$ = water-soluble carbohydrates; TN = total nitrogen; RD = rumen degradability.

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Table 3. Microbiological analysis of the ensiled dairy products after 30 d of ensiling—Experiment 1. Results are given as \log_{10} number of colony forming units g⁻¹.

Treatment ¹	Lactobacilli	Yeasts	Molds	
1	8.5	<2.0	<2.0	
2	8.5	<2.0	<2.0	
3	7.4	<2.0	<2.0	
4	7.0	<2.0	2.9	

¹Treatments: 1) cottage cheese and straw; 2) cottage cheese, straw, and molasses; 3) milk and straw; 4) milk, straw, and molasses.

column (Hewlett Packard, Waldbronn, Germany) over a temperature range of 40 to 230°C. Rumen degradability was measured after 48 h by the in situ procedure of Meherez and Ørskov (1977). Lactobacilli were enumerated via pour-plate technique in Rogosa agar (Oxoid) and yeasts and molds by spread plate on malt extract agar (Oxoid) acidified to pH 4.0 with 10% lactic acid. The plates were incubated at 30°C for 3 d.

Statistical analysis included ANOVA and Duncan's multiple range test using the SAS general linear means procedure (SAS, 1982).

RESULTS

Table 1 gives the analyses of the raw materials for both ensiling experiments. After 1 wk of storage at 4°C, the number of lactobacilli in the milk increased considerably. However, increased lactobacilli numbers did not decrease the pH nor WSC content of the milk; pH even increased slightly, probably due to protein breakdown. Molasses was not analyzed and was assumed to contain 50% WSC (Ashbell et al., 1995).

Table 2 gives the analyses of the resulting dairy products and straw silages after 30 d of ensiling in experiment 1. Mixing the dairy products with the straw was designed to obtain a DM content above 350 g kg⁻¹ according to the DM content of the separate ingredients. The ratio at which cottage cheese and straw were combined in this experiment resulted in somewhat lower DM than expected. The pH of the cottage cheese and



Figure 1. pH change during ensiling of dairy products with straw and molasses.

straw mixture decreased only in the treatment that was augmented with molasses as a carbohydrate source fo: the fermentation. The butyric acid found in the cottage cheese and straw silages may suggest the beginning of clostridial fermentation in that treatment. The milk and straw mixture fermented readily even without molasses due to the relatively high lactose content of the milk, as seen from the decrease in pH and build-up of lactic acid, the major fermentation product. The RD of the cottage cheese and straw silages were higher than straw, but RD of the milk and straw silages were similar to straw. Lactobacilli were the major microbial population, of those measured, in all silages (Table 3). The number of lactobacilli in the cottage cheese mixtures (treatments 1 and 2) were a log cycle higher than in the milk combinations (treatments 3 and 4); yet the pH was higher and the lactic acid content lower in the treatments containing cottage cheese (Table 2). This result is attributed to the cottage cheese being a lactic acid fermentation product, so lactic acid bacteria numbers had increased and WSC had been depleted during its manufacturing. The milk had more WSC (lactos

Table 4. Chemical analysis of the ensiled dairy products after 36 d—Experiment 2. Results are in g kg⁻¹ DM.

Treatment ¹	DM	pН	WSC^3	Lactic acid	Acetic acid	Ethanol	Crude protein	NH ₃ -N%/TN ³	RD^3
1	362	5.3 ^{u2}	0^{e^2}	25^{62}	1012	4	225^{b2}	16 ⁿ²	60162
2	359	4.0%	5°	86 ^a	$4^{\rm b}$	0	282^{a}	4^{d}	646 ^a
3	339	4.0^{6}	$14^{\rm b}$	92^{a}	17^{a}	4	113 ^c	$10^{\rm h}$	480 ^c
-4	338	$3.9^{\rm b}$	41 ^a	97^{a}	$9^{i_{i}}$	Ο	118^{d}	5	490

¹Treatments: 1) cottage cheese and straw; 2) cottage cheese, straw, and molasses; 3) milk and straw; 4) milk, straw, and molasses.

²Within a column, means followed by different superscripts are significantly different (P < 0.05).

³WSC = water-soluble carbohydrates; TN = total nitrogen; RD = rumen degradability.

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Table 5. Numbers of lactobacilli in the ensiled dairy products in Experiment 2. Results are given as \log_{10} number of colony forming units g^{-1} .

Treatment ¹	Day 0	Day 3	Day 7	Day 36 ²
1	6.9	10.0	9.4	8.1
.)	9.6	10.0	9.1	5.5
3	6.0	9.3	9.6	8.6
4	8.7	9.7	8.7	8.9

 1 Treatments: 1) cottage cheese and straw; 2) cottage cheese, straw, and molasses; 3) milk and straw; 4) milk, straw, and molasses.

²On d 36 Treatment 1 had also 3.2 and 5.7 yeasts and molds, respectively.

available for the ensiling fermentation that permitted accumulation of lactic acid and decreased pH.

Figure 1 gives the change in pH during ensiling of the mixtures of dairy products and straw (experiment 2). Except for the cottage cheese and straw treatment, all treatments fermented well, and even by d 3 their pH values were approximately 4.0.

The higher proportion of straw in these mixtures resulted in a higher DM content in experiment 2 (Table 4). Overall, the results of experiment 2 were similar to experiment 1, in that cottage cheese and straw mixtures needed molasses addition to ensile properly. The silages that comprised of cottage cheese mixtures in experiment 2 had a wider lactic acid to acetic acid ratio than in experiment 1. The improved fermentation of these mixtures can be attributed to the higher DM content and to lower buffering capacity with extra straw. Crude protein content in the second experiment was lower than in experiment 1, probably due to protein breakdown during the 1-wk storage, or due to the greater dilution of straw. The percentage of ammonia N (of total N) was generally higher in the silages containing cottage cheese than in silages that included milk; ammonia levels increased in the silages prepared with milk in the second experiment compared to experiment 1. The highest percent of ammonia N of total N was 164, measured in the second experient in the silages prepared with cottage cheese and straw, indicating only moderate proteolysis.

Rumen degradability decreased in all treatments in experiment 2, as compared with experiment 1, even below the value of straw, which we cannot explain. Table 5 gives the microbial counts during the ensiling period of experiment 2. Results indicate that the numbers of lactobacilli were high in all treatments even by the 3rd d of ensiling.

DISCUSSION

Utilization of returned dairy products for cattle feeding could benefit both the environment and the animal industry. The results of the current experiments indi-

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cate that it is possible to stabilize returned dairy products for animal feeding by ensiling. Ensiling moist dairy products along with straw seems a feasible technology for such commodities. A similar technology is being used in the preservation of citrus peels for animal feeding. Due to high yeast populations the moist peels without amendment undergo an ethanolic fermentation with substantial losses (Ashbell et al., 1987). Therefore, an economical way to stabilize the moist peels is to coensile them with dry broiler litter, a common practice on farms in Israel (Ashbell et al., 1995).

Some returned processed dairy products may not contain enough fermentable carbohydrates necessary for ensiling, as was the case with cottage cheese. They may require addition of an inexpensive source of sugars. However, in a commercial operation, various types of returned dairy products may possibly be combined to provide enough carbohydrates for the ensiling without further addition of other sugar sources.

The current study is only the first step towards realization of such technology. More studies are required to test this idea in larger scale. One problem associated with this technology might be the separation of the dairy products from the packaging materials. It could well be that a press that squeezes out the products from the package may be a solution. Utilization of waste milk from large dairy farms or from groups of farms by ensiling with straw for later use as feed seems more feasible at present, as it does not require the separation of products from small containers.

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